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Synthesis, conformation and glycosylation reaction of substituted 2,6-dioxabicyclo[3.1.1]heptanes: 1,3-anhydro-2,4-di-*O*-benzyl-α-L-arabinopyranose

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Abstract

The title 1,3-anhydro sugar (10) was obtained as crystals by the ring closure of 3-O-acetyl-2,4-di-O-benzyl- α , β -L-arabinopyranosyl chloride (9) which was prepared from methyl β -L-arabinopyranoside via a sequence of reactions involving initial selective 3-O-allylation. Methanolysis of 10 gave 1,3-cis arranged methyl 2,4-di-O-benzyl- α -L-arabinopyranoside as the major product. Glycosylation of 10 with 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose or with serine derivatives resulted in the same stereochemical outcome, while glycosylation of the 1,3-anhydro galacto- (19) and manno-pyranose (24) analogues afforded predominantly 1,3-trans ring opening products. Conformational analysis of 10, 19 and 24 revealed that the C-5 head of 10 is flexible. The conformation of 10 in solution is close to E_2 while 19 and 24 prefer the $B_{2.5}$ form.

Keywords: 2,6-Dioxabicyclo[3.1.1]heptanes; Synthesis; Conformation

1. Introduction

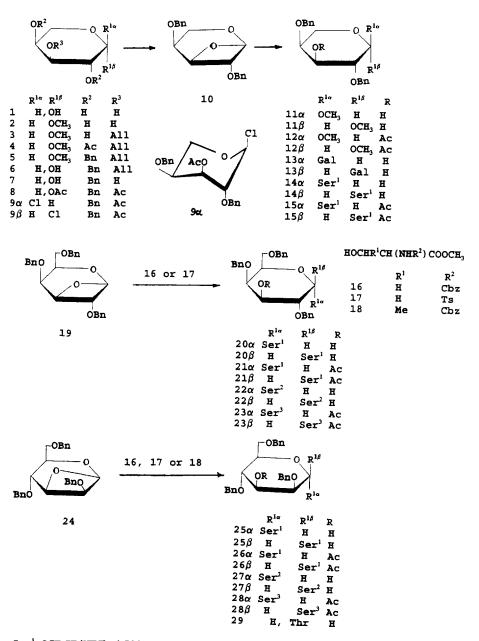
As part of an ongoing research-effort on 2,6-dioxabicyclo[3.1.1]heptane derivatives, we have synthesized 1,3-anhydro- β -L-rhamno-[1], - β -D-galacto-[2], -6-deoxy- β -D-gluco-[3], -6-azido-6-deoxy- β -D-manno-[4], - β -D-fuco-[5], and - β -D-talo-pyanose [6] benzyl ethers. The synthesis of 1,3-anhydro- β -D-gluco-[7,8] and - β -D-manno-pyranose [9,10] analogues had been reported earlier by Schuerch's group. We report here on the

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synthesis, conformation, and glycosylation reaction of 1,3-anhydro-2,4-di-O-benzyl- α -L-arabinose whose stereoregular polymerization could afford 1 \rightarrow 3-linked arabinopyranan, a useful model compound for polysaccharide research.

2. Results and discussion

L-Arabinopyranose (1) was converted to methyl β -L-arabinopyranoside (2) with strongly acidic resin (H+ form) as catalyst [11]. Compound 2 was selectively allylated at C-3 using the method for 3-O-alkylation of methyl α -D-galactopyranoside [2] via a stannylene complex, and methyl 3-O-allyl-\(\beta\)-arabinopyranoside (3) was obtained in satisfactory yield. Removal of most of the tetrabutylammonium iodide used in the monoallylation from the reaction mixture was helpful for the further purification of 3 by column chromatography, and the recovered tetrabutylammonium iodide from ethyl acetate or diethyl ether could be reused. The structure of 3 was confirmed by acetylation to afford methyl 2,4-di-O-acetyl-3-O-allyl-β-L-arabinopyranoside (4) whose ¹H NMR spectrum showed H-2 and H-4 shifted downfield. Benzylation of 3 with benzyl bromide and sodium hydride in oxolane afforded methyl 3-O-allyl-2,4-di-O-benzyl-B-Larabinopyranoside (5). Acid hydrolysis of $5 (\rightarrow 6)$ followed by deallylation with tris(triphenylphosphine)rhodium(I) chloride as the catalyst yielded 2,4-di-O-benzyl-Larabinopyranose (7). Acetylation of 7 with acetic anhydride in pyridine gave the 1.3-diacetate 8. Both compounds 7 and 8 were obtained as a mixture of α and β anomers as indicated by their ¹H NMR spectra. Compound 8 was reacted with hydrogen chloride in diethyl ether to furnish the key intermediates 3-O-acetyl-2,4-di-O-benzyl-α- (9α) and $-\beta$ -L-arabinopyranosyl chloride (9β) , which were separated in pure form by analytical LC, in a ratio of about 1:1. Compound 9α assumes a conformation close to ${}^{1}C_{4}$ as indicated from its ${}^{1}H$ NMR spectrum which yielded small values for ${}^{3}J_{H1.H2}$ (1.1 Hz), ${}^3J_{\rm H2,H3}$ (2.9 Hz), and a relatively large value for ${}^3J_{\rm H4,H5}$ (6.7 Hz), while 9β exists in a 4C_1 form as shown from ${}^3J_{\rm H1,H2}$ (3.7 Hz), a large value of ${}^3J_{\rm H2,H3}$ (10.3 Hz), and a small ${}^3J_{\rm H4,H5}$ (2.0 Hz). Similar conformation flipping was observed for compound 4 ${}^{3}J_{\rm H2.H3} = 2.0$ Hz). An alternative method for the preparation of 3-O-acetyl-2,4-di-Obenzyl-L-arabinopyranosyl chloride was also examined. Deallylation of 5 with palladium dichloride as the catalyst in methanol furnished methyl 2,4-di-O-benzyl-β-Larabinopyranoside (11 β) in good yield. Chlorination of 11 β with hydrogen chloride in 1:1 HOAc-CH₂Cl₂ gave the chloride compound in low yield (40%), together with some by-products, and product purification was difficult. Similar treatment of methyl 3-O-acetyl-2,4-di-O-benzyl- β -L-arabinopyranoside (12 β) was also troublesome, and so the alternative method was not used. Compound 9B contains a trans-oriented C-3-Oacetate as a potential alkoxide and C-1-Cl as a leaving group, fulfilling the requirement for a backside attack in a ring closure reaction, whereas 9α does not have such a relation between C-1 and C-3. However, it was found that treatment of either 9α or 9β with potassium tert-butoxide in anhydrous oxolane yielded 1,3-anhydro-2,4-di-O-benzyl- α -L-arabinopyranose (10), a crystalline compound, as the major product, indicating that isomerization of 9α to 9β occurred easily during the ring closure reaction. This behavior is different from the 1,2-ring closure reaction [12,13] in which the precursor for



Ser¹=OCH₂CH (NHCbz) COOCH₃ Ser²=OCH₂CH (NHTs) COOCH₃ Ser³=OCH₂CH (NACTs) COOCH₃ Thr=OCH (CH₃) CH (NHCbz) COOCH₃ Gal=6-O-1,2:3,4-di-O-isopropylidine-α-D-galactopyranos-6-yl

the ring closure must have C-2-O-acetate and C-1-halide in a trans-relation. As either 9α or 9β was readily converted to the anhydro sugar 10, a practical preparation of 10 was carried out with the mixture of 9α and 9β . The ¹H NMR spectrum of 10 showed a characteristic triplet at δ 5.58 for H-1 with ${}^3J_{\rm H1,H2} = J_{\rm H1,H3} = 3.4$ Hz, a similar pattern to the spectrum of 1,3-anhydro-2,4,6-tri-O-benzyl- β -D-galactopyranose [2] (δ 5.58 for H-1, ${}^{3}J_{\text{H1.H2}} = J_{\text{H1.H3}} = 3.5$ Hz). It was reported that treatment of 3-O-acetyl-2,4,6-tri-O-benzyl- α -D-glucopyranosyl chloride [8] and 3-O-acetyl-2,4-di-O-benzyl-6-deoxy- α -D-glucopyranosyl chloride [3] with potassium tert-butoxide in oxolane yielded the corresponding glucal derivatives as the main products, by trans-diaxial elimination of hydrogen chloride from C-1 and C-2. This behavior was not observed during similar treatment of 3-O-acetyl-2,4,6-tri-O-benzyl-α-D-galactopyranosyl chloride and 3-Oacetyl-2,4-di-O-benzyl-6-deoxy- α -D-galactopyranosyl chloride [5]. In the present study of 3-O-acetyl-2,4-di-O-benzyl-L-arabinopyranosyl chloride treated with potassium tertbutoxide in dry oxolane, 1,3-anhydro sugar was formed as the main product (77%) while the glycal derivative was obtained as a by-product (15%; its ¹H NMR spectrum gave H-1 as a singlet at δ 6.25 ppm).

For investigation of the conformation of 10 calculations by molecular mechanics [14] were carried out. MMX is a new program recently made available [15] for performing molecular mechanics calculations on an IBM 386 or equivalent, and a detailed description of this method of operation has appeared [16]. This program, based on the MM2

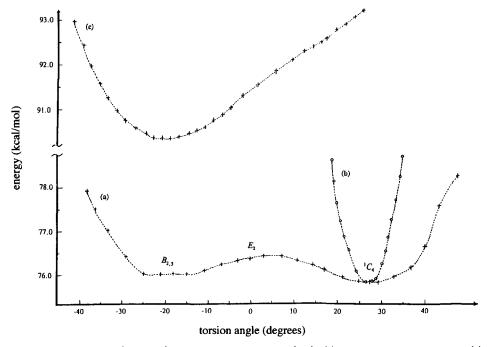


Fig. 1. Plots of energy (kcal/mol) versus the torsion angle (deg). (a) C-3-C-4-C-5-O-5 for 10. (b) C-3-C-2-C-1-O-3 for 10. (c) C-3-C-4-C-5-O-5 for 19.

force field of Allinger, has already gained considerable acceptance [17-19]. Previously we found that the conformational properties of per-O-benzylated 1,2-anhydro-D-talopyranose in the solid state [20], and per-O-benzylated 1,2-anhydro-D-xylopyranose in the solution state [21], could be reproduced satisfactorily using MMX. Here we conducted calculations for description of the conformational properties of 10 and 19 as shown in Fig. 1. In Fig. 1, plot a represents the relationship between total energy and the C-3-C-4-C-5-O-5, torsion angle which indicates the energy changes upon pseudorotation from the $B_{2,5}$ form to the ${}^{1}C_{4}$ form for the pyranose ring of 10. It was found that a conformation close to ${}^{1}C_{4}$ had the lowest energy (75.82 kcal/mol), and a conformation close to $B_{2.5}$ had a similar energy (76.00 kcal/mol), while the E_2 form had an energy of 76.40 kcal/mol. When the C-3-C-4-C-5-O-5 torsion angle changed over a wide range (-25°) to $+37^{\circ}$, accompanied by a symmetrical change in the C-4-C-5-O-5-C-1 torsion angle) the energy fluctuation was relatively small (75.82-76.42 kcal/mol), thus revealing that the C-4-C-5-O-5 moiety (C-5 head) of the pyranose ring is flexible. Plot b describes the rigidity of the four membered ring part in 10. A sharp increment of the total energy was observed when the C-3-C-2-C-1-O-3 torsion angle assumed a value outside the 29°-25° range, indicating that the four membered ring is considerably rigid. Plot c indicates the conformational properties of 19, which are different from those of 10. There is only one energy well for the C-3-C-4-C-5-O-5 torsion angle at about 19°, corresponding to the $B_{2.5}$ conformation; the E_2 and 1C_4 forms had 1 kcal/mol and 2.8 kcal/mol higher energies, respectively, than that of $B_{2.5}$. The calculated torsional angles and coupling constants for the conformations corresponding to ${}^{1}C_{4}$, E_{2} , and $B_{2,5}$ of 10, and also for the conformation (Exp) having coupling constants close to the experimental values, are shown in Table 1. We suggest from the data that the actual conformation of 10 in the solution state is close to E_2 . For comparison, the reproduced conformations, being close to $B_{2.5}$ for 19 and 24 calculated according to the observed coupling constants, are also shown in Table 1 (19 Exp, 24 Exp).

Methanolysis of the 1,3-anhydropyranose 10 was conducted to investigate its reactivity. No reaction occurred when a methanol solution of 10 was stirred at room temperature for 16 h. However, when freshly fused ZnCl₂ was added as the catalyst, all of the starting material disappeared and two compounds formed in a ratio of 3.5:1 as indicated by analytical LC. The major product was identified by ¹H NMR as methyl 2,4-di-O-benzyl- α -L-arabinopyranoside (11 α), and the minor one was methyl 2,4-di-Obenzyl- β -L-arabinopyranoside (11 β) which was identical to the product obtained from deallylation of 5. Thus, an unexpected ring opening of the 1,3-anhydro sugar produced mainly the 1,3-cis glycoside. Further study of the glycosidic coupling reaction of 10 with 1,2:3,4-di-O-isopropylidene- α -D-galactopyranose as the acceptor gave a similar result, i.e. α -linked disaccharide 13 α was the major product. The 4C_1 conformation and α (1,2-trans) anomeric configuration of the arabinose moiety of 13α were established from its ¹H NMR spectrum, which yielded an upfield shift for H-1' (8 4.30, axial H), and large values for ${}^3J_{\rm H1',H2'}$ (8.3 Hz) and ${}^3J_{\rm H2',H3'}$ (9.2 Hz). The structural assignment of 13 β was also achieved from its ¹H NMR spectrum, which yielded a downfield H-1' (δ 4.89, equatorial H), a small ³ $J_{\rm H1',H2'}$ (3.8 Hz), and a large ³ $J_{\rm H2',H3'}$ (9.0 Hz, ⁴ C_1 form). The structure of 14 α was assigned by the same method (δ 4.38 for H-1, α -linkage; $^3J_{\rm H_1\,H_2} = 5.4$ Hz, $^3J_{\rm H_2\,H_3} = 7.1$ Hz for a 4C_1 conformation), and the assignment was

Table 1 Selected torsion angles and ¹H-¹H vicinal coupling constants obtained by MMX for 10, 19, and 24

Angle	Magnitude (°)					
	10				19	24
	C_4	B _{2,5}	E_2	Exp	Exp	Exp
O-5-C-1-C-2-C-3	-87.6	-82.6	-87.8	-87.3	-80.2	-86.4
C-1-C-2-C-3-C-4	82.5	79.2	83.4	83.3	77.0	83.2
C-2-C-3-C-4-C-5	-63.9	-34.6	-50.5	-49.3	-32.3	-38.2
C-3-C-4-C-5-O-5	27.8	-28.8	0	-3.0	-21.8	-35.3
C-4-C-5-O-5-C-1	-31.1	29.9	-0.6	3.1	23.6	37.4
C-5-O-5-C-1-C-2	71.5	34.2	53.5	51.0	30.5	35.5
C-5-O-5-C-1-O-3	-28.5	-64.9	-47.5	-50.0		
O-5-C-1-O-3-C-3	85.3	88.3	88.6	88.9		
C-1-O-3-C-3-C-4	-84.1	-85.2	-85.6	-85.4		
O-3-C-3-C-4-C-5	33.5	61.9	47.4	48.6		
C-1-O-3-C-3-C-2	28.5	30.0	27.2	27.4		
O-3-C-3-C-2-C-1	-26.5	-27.9	-25.4	-25.6		
C-3-C-2-C-1-O-3	26.6	28.0	25.5	25.7		
C-2-C-1-O-3-C-3	-28.5	-30.0	-27.3	-27.5		
Coupling constants (H	z, modified Ka	rplus) for ind	icated torsion a	angles (°)		
$J_{1,2} / \phi_{1,2}$	5.5 / 31	5.5 / 31	5.6 / 30	5.6 / 30	5.6 / 29	1.1 / 101
$J_{2,3} / \phi_{2,3}$	8.2 / 26	8.0 / 28	8.2 / 26	8.2 / 26	7.6 / 33	1.6 / 106
$J_{3,4} / \phi_{3,4}$	0.9 / 79	2.9 / 55	1.9 / 65	1.9 / 64	3.8 / 48	3.6 / 62
$J_{4.5} / \phi_{4.5}$	8.9 / 29	5.6 / 24	8.8 / 1	8.5 / 1	1.8 / 49	6.6 / 145
$J_{4.5'} / \phi_{4.5'}$	8.4 / 148	1.5 / 92	3.8 / 118	3.4 / 115		
Observed coupling co	nstants (Hz) for	r 10, 19, and	24			
				10	19	24
$J_{1,2}$				3.4	3.5	0
$J_{2,3}^{1,2}$				6.1	5.5	0
$J_{3,4}^{2,3}$				1.2	2.3	3.4
$J_{4,5}^{3,4}$				7.2	1.2	6.6
$J_{4,5'}^{7,5}$				3.4	_	_

further confirmed from its acetylated derivative 15α (δ 4.40 for H-1, ${}^3J_{\rm H1,H2}=5.1$ Hz). The 1H NMR of 14β was not easily assigned due to overlap of some signals, whereas its acetylated derivative 15β gave a clear spectrum (δ 4.80 for H-1, β -linkage; ${}^3J_{\rm H1,H2}=4.0$ Hz, ${}^3J_{\rm H2,H3}=11.8$ Hz for the 4C_1 form). The stereochemical outcome for the glycosylation of 10 slightly changed as the reaction temperature was varied ($\alpha:\beta=2:1$ at -10° C, and 3:1 at 30° C). It was also found that, with boron trifluoride etherate or silver triflate as the catalyst at room temperature, both the stereochemical outcome and the total yield were somewhat different (BF₃Et₂O, $\alpha:\beta=1.5:1$, yield 63%; ZnCl₂, $\alpha:\beta=3:1$, total yield 84%; AgOTf, $\alpha:\beta=3.2:1$, yield 88%).

To rationalize the 1,3-cis ring opening of 10, further studies on the glycosylation of 10, 1,3-anhydro-2,4,6-tri-O-benzyl- β -D-galacto- (19) [2], and - β -D-mannopyranose (24) (prepared from methyl α -D-mannopyranoside through selective 3-O-allylation, then benzylation followed by the same procedures as reported in [10]) with N-benzyl-oxycarbonyl- (16) and N-tosyl-L-serine (17) or N-benzyloxycarbonyl-L-threonine (18)

methyl ester derivatives were carried out. Compounds 19 and 24 gave normal 1,3-trans glycopeptides predominantly, while 10 still gave the 1,3-cis arranged product as the major one. The assignment of anomeric configuration of mannopyranosyl derivatives was based upon the chemical shift of H-1, i.e. the α anomers gave an H-1 signal at downfield (δ 4.85 for 25 α , δ 4.81 for 27 α) whereas the β anomers gave an H-1 signal upfield (δ 4.47 for 25 β , δ 4.38 for 27 β). The difference in the chemical shift of H-1 between 25 α and 25 β (0.38 ppm), or between 27 α and 27 β (0.43 ppm) is close to that (0.46 ppm) between allyl 3-O-allyl-2,4,6-tri-O-benzyl- α -D-mannopyranoside (δ 4.92 for H-1) [22] and allyl 3,4,6-tri-O-benzyl- β -D-mannopyranoside (δ 4.46 for H-1) [23]. The detailed mechanism for the ring opening of 1,3-anhydro sugars remains to be elucidated.

3. Experimental

General methods.—Melting points were determined with a "Mel-Temp" apparatus. Optical rotations were determined with a Perkin-Elmer model 241-MC automatic polarimeter for solutions in a 1-dm, jacketed cell. ¹H and ¹³C NMR spectra were recorded with Varian XL-400 and Varian XL-200 spectrometers, for solutions in CDCl₃ with tetramethylsilane (Me₄Si) as the internal standard. Chemical shifts are expressed in ppm downfield from the internal Me₄Si absorption. Mass spectra were recorded with a JDS-D 3005 mass spectrometer using a direct-insertion technique to introduce the sample. Analytical LC was performed with a pump (model YSB-2, made in China), stainless-steel columns packed with silica gel (10×150 mm, or 4.6×250 mm) or Lichrosorb-NH₂ (4.6 × 250 mm), a differential refractometer (Perkin-Elmer LC-25 RI Detector), and ethyl acetate-petroleum ether (bp 60-90°C) as the eluent at a flow rate of 1-4 mL/min. Thin-layer chromatography (TLC) was performed on silica gel HF, detection being affected by charring with 30% (v/v) sulfuric acid in methanol or sometimes by UV detection. Column chromatography was conducted by elution of a column (16×240 mm, 18×300 mm, 35×400 mm) of silica gel (100-200 mesh). Solutions were concentrated at a temperature < 60°C under diminished pressure.

Molecular mechanics calculations [14] were carried out using the MMX program [15] embedded in PCMODEL-386 on an AST-386 computer. The dielectric constant used throughout the calculations was 1.5. Each calculated total energy consisted of stretching, bending, stretching-bending, torsional, van der Waals, and dipole-dipole contributions. Calculations of each individual conformation were carried out with one fixed torsion angle (C-3-C-4-C-5-O-5 for evaluation of the flexibility of the pyranose moiety C-4-C-5-O-5 of 10 and 19; and C-3-C-2-C-1-O-3 for evaluation of the rigidity of the four membered ring C-3-C-2-C-1-O-3 of 10). Calculations with the two torsion angles H-3-C-3-C-4-H-4 ($\Phi_{3,4}$) and H-4-C-4-C-5-H-5 ($\Phi_{4,5}$) fixed at values correlated to the observed coupling constants $^3J_{\rm H3,H4}$ and $^3J_{\rm H4,H5}$ gave the deduced conformation for 19 and 24 (Exp in Table 1).

Methyl 3-O-allyl- β -L-arabinopyranoside (3).—A mixture of methyl β -L-arabinopyranoside (2) [11] (3.28 g, 20 mmol) and dibutyltin oxide (5.5 g, 22.0 mmol) in absolute methanol (160 mL) was boiled under reflux. After the mixture became transparent, heating was continued for 2 h, and the solution was concentrated to give a

white foamy residue (8.8 g). Tetrabutylammonium iodide (7.4 g, 20 mmol), allyl bromide (20.4 mL, 240 mmol) and toluene (200 mL) were added to the residue and the mixture was stirred for 1 day at 60°C. TLC (EtOAc) showed the presence of major product 3 together with small amounts of the di-O-allylated compound and methyl 4-O-allyl- β -L-arabinopyranoside as by-products. After evaporation of the solvent, most of the tetrabutylammonium iodide was recovered from the residue by precipitation with ethyl acetate or diethyl ether. The brownish residue was purified by chromatography on a column of silica gel (1:1 petroleum ether-EtOAc) to afford yellowish syrupy 3 (2.81 g, 69%); ¹H NMR: δ 6.00–5.70 (m, 1 H, CH₂=CH-CH₂), 5.25–5.15 (m, 2 H, $CH_2 = CH - CH_2 -)$, 4.85 (d, 1 H, $J_{1,2}$ 3.4 Hz, H-1), 4.72 (dd, 1 H, $J_{1,2}$ 3.4, $J_{2,3}$ 2.8 Hz, H-2), 4.45–4.38 (m, 3 H, H-4 and $CH_4 = CH - CH_2 - J$, 3.98 (dd, 1 H, $J_{2,3}$ 2.8, $J_{3,4}$ 4.1 Hz, H-3), 3.95-3.87 (m, 2 H, H-5,5'), 3.40 (s, 3 H, OC H_3). Methyl 4-O-allyl- β -Larabinopyranoside was the minor product and not isolated in a pure form; its ¹H NMR gave OCH_3 as a singlet at δ 3.38, and its ¹³C NMR gave OCH_3 at δ 55.10. After acetylation, the ¹H NMR spectrum of methyl 2,3-di-O-acetyl-4-O-allyl-β-Larabinopyranoside showed two C H_3 CO signals at δ 2.09 and 2.08.

Compound **3** was acetylated by standard methods and compound **4** was obtained in a quantitative yield as a syrup; $[\alpha]_D$ +181° (c 4.7, CHCl₃); ¹H NMR: δ 5.95–5.70 (m, 1 H, CH₂=CH-CH₂), 5.30 (dd, 1 H, $J_{1,2}$ 3.7, $J_{2,3}$ 2.0 Hz, H-2), 5.25–5.17 (m, 1 H, H-4), 5.15–5.05 (m, 2 H, CH₂=CH-CH₂), 4.91 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1), 4.10–4.00 (m, 2 H, CH₂=CH-CH₂-), 3.87 (dd, 1 H, $J_{2,3}$ 2.0, $J_{3,4}$ 4.2 Hz, H-3), 3.82–8.67 (m, 2 H, H-5,5'), 3.39 (s, 3 H, OCH₃), 2.15, 2.12 (2 s, 6 H, CH₃CO); ¹³C NMR: δ 170.0 (CH₃CO), 134.2 (CH₂=CH-CH₂), 116.6 (CH₂=CH-CH₂-), 96.8 (C-1), 72.0, 70.4, 70.0 (C-3,2, and 4), 68.1 (CH₂=CH-CH₂-), 60.2 (C-5), 55.2 (OCH₃) Anal. Calcd for C₁₃H₂₀O₇: C, 54.17; H, 6.94. Found: C, 54.28; H, 6.89.

Methyl 3-O-allyl-2,4-di-O-benzyl-β-L-arabinopyranoside (5).—To a solution of 3 (2.5 g, 11.0 mmol) in dry oxolane (50 mL) cooled in an ice bath was added sodium hydride (80% in oil; 1.05 g, 35 mmol) with stirring. Benzyl bromide (2.93 mL, 24.2 mmol) was added dropwise to the mixture and the mixture was boiled under reflux with vigorous agitation for 4 h. TLC (4:1 petroleum ether–EtOAc) indicated that the reaction was complete. The remaining sodium hydride was filtered off and the filtrate was concentrated. The residue was partitioned between water and dichloromethane, and the organic layer was dried (Na₂SO₄) and concentrated. Column chromatography (5:1 petroleum ether–EtOAc) gave pure compound 5 as a yellowish syrup (4.5 g, 95%); [α]_D +57.5° (c 3.8, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 10 H, Ph), 6.02–5.82 (m, 1 H, CH₂=CH–CH₂–), 5.38–5.12 (m, 2 H, CH₂=CH–CH₂–), 4.82, 4.67 (2 d, 2 H, J 12.2 Hz, PhCH₂), 4.74, 4.73 (2 d, 2 H, J 10.0 Hz, PhCH₂), 4.64 (d, 1 H, J_{1,2} 3.2 Hz, H-1), 4.20–4.10 (m, 2 H, H-2,4), 3.80–3.55 (m, 3 H, H-3,5, and 5'), 3.39 (s, 3 H, OCH₃). Anal. Calcd for C₂₃H₂₈O₅: C, 71.87; H, 7.29. Found: C, 71.66; H, 7.32.

3-O-Allyl-2,4-di-O-benzyl-L-arabinopyranose (6).—A mixture of 5 (4.0 g, 10.4 mmol), acetic acid (80%, 24 mL) and hydrochloric acid (1 N, 6 mL) was boiled under reflux for 2 h, at the end of which time TLC (2:1 petroleum ether-EtOAc) showed that all of the starting material had disappeared. Solid sodium bicarbonate was added to neutralize the mixture. The mixture was extracted with CH_2Cl_2 (3 × 20 mL), the extract was washed with saturated sodium bicarbonate and then water, dried (Na₂SO₄) and

concentrated to a syrup. Purification of the syrup by column chromatography (2:1 petroleum ether–EtOAc) yielded crystalline **6** consisting of β and α anomers in a ratio of 2.5:1 (3.18 g, 82.7%); mp 32–34°C, [α]_D +25° (c 4.7, CHCl₃); ¹³C NMR: δ 134.2 (CH₂=CHI–CH₂- of β anomer), 134.0 (CH₂=CH–CH₂ of α anomer), 128.1, 127.7 (Ph–C of β anomer), 127.8, 127.6 (Ph–C of α anomer), 116.6 (CH₂=CH–CH₂- of α anomer), 116.2 (CH₂=CH–CH₂- of β anomer), 94.2 (C-1 of α anomer), 91.8 (C-1 of β anomer), 77.0, 76.1, 75.8, 73.2, and 71.9 (2 PhCH₂, C-2,3, and 4 of β anomer), 77.8, 76.9, 76.2, 72.8, and 71.6 (2 PhCH₂, C-2,3, and 4 of α anomer), 71.4 (CH₂=CH–CH₂- of α anomer), 71.1 (CH₂=CH–CH₂- of β anomer), 60.4 (C-5 of α anomer), 59.5 (C-5 of α anomer). Anal. Calcd for C₂₂H₂₆O₅ · 0.5H₂O: C, 69.65; H, 7.12. Found: C, 69.78; H, 7.16.

2,4-Di-O-benzyl-L-arabinopyranose (7).—Compound 6 (2.04 g, 5.5 mmol) was dissolved in ethanol (90%, 50 mL) and tris(triphenylphosphine)rhodium(I) chloride (100 mg, 0.11 mmol) was added to the solution. The mixture was refluxed and the reaction was monitored by TLC (1:1 petroleum ether–EtOAc) until all of the starting material was consumed (about 5 h). The mixture was filtered and the filtrate was concentrated to give a yellowish solid; recrystallization from EtOAc–petroleum ether afforded 7 as white crystals consisting of β and α anomers in a ratio of 1.5:1 (1.53 g, 84%); mp 104–105.5°C; $[\alpha]_D$ +39° (c 0.6, CHCl₃); ¹H NMR: δ 4.60 (d, H-1 of α anomer), 5.16 (d, H-1 of β anomer). Anal. Calcd for C₁₉H₂₂O₅: C, 69.09; H, 6.67. Found: C, 69.16; H, 6.66.

1,3-Di-O-acetyl-2,4-di-O-benzyl-L-arabinopyranose (8).—Compound 7 (1.06 g, 3.2 mmol) was treated with acetic anhydride (2 mL) in pyridine (3 mL) by a standard method. Compound 8 was obtained in quantitative yield as a syrupy mixture of α and β anomers in a ratio of 2:1; $[\alpha]_D + 64^\circ$ (c 2.7, CHCl₃); ¹H NMR: δ 7.38–7.18 (m, 10 H, Ph), 6.35 (d, 0.33 H, $J_{1,2}$ 3.4 Hz, H-1 of β anomer), 5.68 (d, 0.66 H, $J_{1,2}$ 5.6 Hz, H-1 of α anomer), 5.15 (dd, 0.33 H, $J_{1,2}$ 3.4, $J_{2,3}$ 6.2 Hz, H-3 of β anomer), 5.05 (dd, 0.66 H, $J_{1,2}$ 5.6, $J_{2,3}$ 10.2 Hz, H-3 of α anomer), 4.70–4.43 (m, 4 H, 2 PhC H_2), 4.18–3.58 (m, 4 H, H-2,4,5, and 5'), 2.10, 2.04 (2 s, 1.98 H, 2 C H_3 CO of β anomer), 2.09, 2.01 (2 s, 3.76 H, 2 C H_3 CO of α anomer). Anal. Calcd for $C_{23}H_{26}O_7$: C, 66.67; H, 6.28. Found: C, 66.69; H, 6.20.

Compound **8** was also prepared by acetolysis of methyl 2,4-di-O-benzyl- β -L-arabinopyranoside (11 β). Compound 11 β (170 mg, 0.49 mmol) was treated with acetic anhydride (3 mL) containing acetic acid (1 mL) and concentrated H₂SO₄ (0.03 mL) for 1.5 h. The reaction mixture was neutralized with ice-cooled aqueous potassium carbonate and extracted with CH₂Cl₂. The organic layer was concentrated to a syrup, which was purified by column chromatography (with 4:1 petroleum ether–EtOAc as the eluent) to give syrupy **8** (96 mg, 47%, α : β = 2:3).

3-O-Acetyl-2,4-di-O-benzyl- α - (9α) and - β -L-arabinopyranosyl chloride (9β).—A solution of compound 8 (400 mg, 0.97 mmol) in dry diethyl ether (35 mL) was saturated with hydrogen chloride gas under a nitrogen atmosphere at 0°C and the solution was kept at room temperature in a sealed bottle for 1 h. TLC (3:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solution was concentrated to a syrup, which was dissolved in CH₂Cl₂ (1 mL) and the solvent evaporated. This procedure was repeated 6 times. The product was then purified by analytical LC (5:1 petroleum

ether–EtOAc) to give the chlorides $\mathbf{9}\alpha$ and $\mathbf{9}\beta$ in a ratio of about 1:1 with a total yield of 91%; For $\mathbf{9}\alpha$, $[\alpha]_{\rm D}$ +68.8° (c 2.2, CHCl₃); ¹H NMR: δ 7.41–7.22 (m, 10 H, Ph), 5.80 (d, 1 H, $J_{1,2}$ 1.1 Hz, H-1), 5.26 (t, 1 H, $J_{2,3} = J_{3,4} = 2.9$ Hz, H-3), 4.65, 4.64 (2 d, 2 H, J 12.0 Hz, PhC H_2), 4.59, 4.51 (2 d, 2 H, J 11.8 Hz, PhC H_2), 4.11 (d, 1 H, $J_{4,5}$ 0, $J_{5,5'}$ 12.5 Hz, H-5), 3.98 (dd, 1 H, $J_{1,2}$ 1.1, $J_{2,3}$ 2.9 Hz, H-2), 3.93 (dd, 1 H, $J_{3,4}$ 2.9, $J_{4,5}$ 6.7 Hz, H-4), 3.68 (dd, 1 H, $J_{4,5'}$ 6.7, $J_{5,5'}$ 12.5 Hz, H-5), 2.13 (s, 3 H, C H_3 CO). Anal. Calcd for C₂₁H₂₃ClO₅: C, 64.53; H, 5.89. Found: C, 64.48; H, 6.06. For $\mathbf{9}\beta$, $[\alpha]_{\rm D}$ +151.8° (c 2.5, CHCl₃). ¹H NMR: δ 7.37–7.22 (m, 10 H, Ph), 6.14 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1), 5.21 (dd, 1 H, $J_{2,3}$ 10.3, $J_{3,4}$ 3.2 Hz, H-3), 4.69, 4.68 (2 d, 2 H, J 12.9 Hz, PhC H_2), 4.62, 4.53 (2 d, 2 H, J 12.3 Hz, PhC H_2), 4.19 (dd, 1 H, $J_{1,2}$ 3.7, $J_{2,3}$ 10.3 Hz, H-2), 4.06 (dd, 1 H, $J_{4,5}$ 0.9, $J_{5,5}$ 12.7 Hz, H-5), 3.98–3.93 (m, 1 H, H-4), 3.86 (dd, 1 H, $J_{4,5}$ 2.0, $J_{5,5}$ 12.7 Hz, H-5'), 2.02 (s, 3 H, CH_3 CO).

1,3-Anhydro-2,4-di-O-benzyl-α-L-arabinopyranose (10).—To a 1:1 mixture of 9α and 9β (101 mg, 0.26 mmol) in dry oxolane (5 mL) was added potassium tert-butoxide (90 mg, 0.8 mmol), and the mixture was stirred at room temperature for 2 h, at the end of which time TLC (3:1 petroleum ether–EtOAc) indicated that the starting material had disappeared. The mixture was concentrated to dryness, and the residue was repeatedly extracted with 3:1 petroleum ether–EtOAc. Concentration of the combined extracts yielded 10 as a syrup, which crystallized upon standing in a refrigerator for several days (62.2 mg, 77%); mp 31°C; [α]_D +25° (c 1, CHCl₃); ¹H NMR: δ 7.40–7.22 (m, 10 H, Ph), 5.58 (t, 1 H, $J_{1,2} = J_{1,3} = 3.4$ Hz, H-1), 4.85–4.72 (m, 1 H, $J_{1,3} = 3.4$, $J_{2,3} = 3.4$, J

Methyl 2,4-di-O-benzyl- α - (11 α) and - β -L-arabinopyranoside (11 β).—Compound 10 (50 mg, 0.16 mmol) was dissolved in anhyd MeOH (5.0 mL) in the presence of freshly fused ZnCl₂ (23.0 mg, 0.17 mmol) and the solution was kept for 1 h at room temperature. TLC (2:1 petroleum ether-EtOAc) indicated that the reaction was complete. The solvent was evaporated and water (6 mL) was added to the residue. The mixture was extracted with CH_2Cl_2 (3 × 2 mL), and the combined extracts were dried (Na₂SO₄), and concentrated. Analytical LC (3:1 petroleum ether-EtOAc) of the syrup gave 11α (37.7 mg, 68.4%) and 11β (10.7 mg, 19.6%); For 11α , H NMR: δ 7.40-7.20 (m, 10 H, Ph), 4.76, 4.60 (2 d, 2 H, J 11.7 Hz, PhC H_2), 4.68, 4.62 (2 d, 2 H, J 12.0 Hz, PhC H_2), 4.39 (d, 1 H, $J_{1,2}$ 4.4 Hz, H-1), 3.95 (dd, 1 H, $J_{4,5}$ 6.4, $J_{5,5'}$ 12.0 Hz, H-5), 3.89 (dd, 1 H, $J_{2.3}$ 6.6, $J_{3.4}$ 3.4 Hz, H-3), 3.78 (ddd, 1 H, $J_{3.4}$ 3.4, $J_{4.5}$ 6.4, $J_{4,5'}$ 2.9 Hz, H-4), 3.59 (dd, 1 H, $J_{1,2}$ 4.4, $J_{2,3}$ 6.6 Hz, H-2), 3.48 (dd, 1 H, $J_{4,5'}$ 2.9, $J_{5,5'}$ 12.0 Hz, H-5'), 3.47 (s, 3 H, OC H_3), 2.40 (bs, 1 H, OH). Acetylation of 11 α gave 12 α as a syrup; $[\alpha]_D + 42.5^\circ$ (c 0.6, CHCl₃); ¹H NMR: δ 7.40–7.22 (m, 10 H, Ph), 4.95 (dd, 1 H, $J_{2,3}$ 8.5, $J_{3,4}$ 3.4 Hz, H-3), 4.82, 4.52 (2 d, 2 H, J 11.7 Hz, PhC H_2), 4.64, 4.62 (2 d, 2 H, J 12.2 Hz, PhCH₂), 4.35 (d, 1 H, J_{1,2} 6.1 Hz, H-1), 4.10 (dd, 1 H, J_{4.5} 4.2, $J_{5,5'}$ 12.5 Hz, H-5), 3.82 (ddd, 1 H, $J_{3,4}$ 3.4, $J_{4,5}$ 4.2, $J_{4,5'}$ 2.0 Hz, H-4), 3.76 (dd, 1 H, $J_{1,2}$ 6.1, $J_{2,3}$ 8.5 Hz, H-2), 3.52 (s, 3 H, OC H_3), 3.44 (dd, 1 H, $J_{4,5'}$ 2.0, $J_{5,5'}$ 12.5 Hz, H-5'), 2.05 (s, 3 H, C H_3 CO). Anal. Calcd for $C_{22}H_{26}O_6$: C, 68.39; H, 6.74. Found:

C, 68.21; H, 6.73. For $\mathbf{11}\boldsymbol{\beta}$, $^1\mathrm{H}$ NMR: δ 7.38–7.25 (m, 10 H, Ph), 4.82–4.59 (m, 5 H, H-1, 2 PhC H_2), 4.03 (dd, 1 H, H-2), 3.82–3.74 (m, 2 H, H-3,4), 3.72–3.66 (m, 2 H, H-5,5'), 3.38 (s, 3 H, OC H_3), 2.18 (bs, 1 H, OH). Acetylation of $\mathbf{11}\boldsymbol{\beta}$ gave $\mathbf{12}\boldsymbol{\beta}$ as a syrup; $[\alpha]_{\mathrm{D}}$ +112° (c 4.6, CHCl $_3$); $^1\mathrm{H}$ NMR: δ 7.40–7.20 (m, 10 H, Ph), 5.20 (dd, 1 H, $J_{2,3}$ 10.3, $J_{3,4}$ 3.4 Hz, H-3), 4.72, 4.52 (2 d, 2 H, J 12.2 Hz, PhC H_2), 4.70 (d, 1 H, $J_{1,2}$ 3.1 Hz, H-1), 4.63, 4.62 (2 d, 2 H, J 12.5 Hz, PhC H_2), 4.00 (dd, 1 H, $J_{1,2}$ 3.1, $J_{2,3}$ 10.3 Hz, H-2), 3.93–3.86 (m, 1 H, H-4), 3.73 (dd, 1 H, $J_{4,5}$ 1.2, $J_{5,5'}$ 12.0 Hz, H-5), 3.64 (dd, 1 H, $J_{4,5}$ 2.2, $J_{5,5'}$ 12.0 Hz, H-5'), 3.41 (s, 3 H, OC H_3), 2.05 (s, 3 H, C H_3 CO).

O-(2,4-Di-O-benzyl- α -L-arabinopyranosyl)- $(1 \rightarrow 6)$ -1,2:3,4-di-O-isopropylidene- α -Dgalactopyranose (13 α) and O-(2,4-di-O-benzyl- β -L-arabinopyranosyl)-(1 \rightarrow 6)-1,2:3,4di-O-isopropylidene- α -D-galactopyranose (13 β).—The 1,3-anhydro sugar 10 (50 mg, 0.16 mmol) was dissolved in anhydrous oxolane (5.0 mL) containing freshly fused ZnCl₂ (20 mg) as the promoter. To the mixture was added a solution of 1,2:3,4-di-Oisopropylidene-α-D-galactopyranose (70 mg, 0.27 mmol) in oxolane (2 mL) in one portion. The mixture was stirred at room temperature for 16 h, at which time TLC (2:1 petroleum ether-EtOAc) indicated that 10 disappeared. The solution was concentrated to a syrup which was subjected to separation by analytical LC with 2:1 petroleum ether-EtOAc as eluent. Compound 13α was obtained as a syrup (57.8 mg, 63.0%); $[\alpha]_{D}$ – 53° (c 0.4, CHCl₃); ¹H NMR δ 7.40–7.20 (m, 10 H, Ph), 5.54 (d, 1 H, $J_{1,2}$ 5.1 Hz, H-1), 4.80, 4.65 (2 d, 2 H, J 11.8 Hz, PhC H_2), 4.67, 4.66 (2 d, 2 H, J 12.6 Hz, PhC H_2), 4.61 (dd, 1 H, $J_{2.3}$ 2.3, $J_{3.4}$ 7.8 Hz, H-3), 4.30 (dd, 1 H, $J_{1.2}$ 5.1, $J_{2.3}$ 2.3 Hz, H-2), 4.30 (d, 1 H, $J_{1',2'}$ 8.3 Hz, H-1'), 4.23 (dd, 1 H, $J_{3,4}$ 8.1, $J_{4,5}$ 1.7 Hz, H-4), 4.20 (dd, $J_{1',2'}$ 8.3 Hz, $J_{2',3'}$ 9.2 Hz, H-2'), 4.16-4.00 (m, 3 H, H-5, 6a, 6b), 3.75 (dd, 1 H, $J_{4'.5'}$ 9.2 Hz, $J_{5'.5''}$ 12.5 Hz, H-5'), 3.70–3.65 (m, 1 H, H-4'), 3.45–3.38 (m, 1 H, H-3'), 3.35 (dd, 1 H, $J_{4'5''}$ 1.8 Hz, $J_{5'5''}$ 12.5 Hz, H-5"), 1.80 (bs, 1 H, OH), 1.52, 1.44, 1.35, 1.34 (4 s, 12 H, 4 C H_3). Anal. Calcd for $C_{31}H_{40}O_{10}$: C, 65.05; H, 6.99. Found: C, 65.14; H, 7.23. Compound 13 β was obtained as a syrup (19.3 mg, 21.1%); [α]_D -8.2° (c 0.1, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 10 H, Ph), 5.53 (d, 1 H, $J_{1,2}$ 5.0 Hz, H-1), 4.89 (d, 1 H, $J_{1',2'}$ 3.8 Hz, H-1'), 4.79, 4.64 (2 d, 2 H, J 11.6 Hz, PhC H_2), 4.63, 4.62 (2 d, 2 H, J 12.8 Hz, PhC H_2), 4.61 (dd, 1 H, $J_{2,3}$ 2.4, $J_{3,4}$ 7.8 Hz, H-3), 4.30 (dd, 1 H, $J_{1,2}$ 5.0, $J_{2,3}$ 2.4 Hz, H-2), 4.22 (dd, 1 H, $J_{3,4}$ 7.8 Hz, $J_{4,5}$ 1.9 Hz, H-4), 4.20 (dd, 1 H, $J_{1',2'}$ 3.8, $J_{2',3'}$ 9.0 Hz, H-2'), 4.09–3.70 (m, 7 H, H-5,6a,6b,3',4',5' and 5"), 2.03 (bs, 1) H, OH), 1.55, 1.46, 1.35, 1.35 (4 s, 12 H, 4 C H_3).

General procedure for glycopeptide preparation.—Amino acid methyl ester (1.2 equiv) was dissolved in dry CH_2Cl_2 (10 mL) in the presence of powdered 4 Å molecular sieves and the mixture was stirred for 10 min. Freshly fused $ZnCl_2$ (1 equiv) and 1,3-anhydo sugar (1 equiv) were then added with vigorous stirring. After 2 h the mixture was filtered and the filtrate was washed with water (3 × 10 mL), dried with Na_2SO_4 , and evaporated to dryness. The crude product was purified by analytical LC on silica gel with 2:1 petroleum ether–EtOAc as the eluent. The following products were obtained starting from 100 mg of 1,3-anhydro sugars 10, 19 and 24 respectively.

O-(2,4-Di-O-benzyl- α - (14α) and - β -L-arabinopyranosyl)-N-benzyloxycarbonyl-L-serine methyl ester (14β) .—Pure 14α (122.1 mg, 70%) and 14β (52.9 mg, 20%) were obtained in a ratio of 3.5:1; For 14α , ¹H NMR: δ 7.40–7.20 (m, 15 H, Ph), 5.71 (d, 1

H, J 8.1 Hz, NH), 5.12 (s, 2 H, PhC H_2 OCO), 4.73, 4.56 (2 d, 2 H, J 11.5 Hz, PhC H_2), 4.69, 4.63 (2 d, 2 H, J 10.8 Hz, PhC H_2), 4.58–4.50 (m, 1 H, CH $_2$ CH), 4.38 (d, 1 H, $J_{1,2}$ 5.4 Hz, H-1), 4.27, 3.89 (2 dd, 2 H, J 10.3, $J_{\text{CH',CH}}$ 3.2 Hz, $J_{\text{CH'',CH}}$ 4.7 Hz, C H_2 CH), 3.80 (s, 3 H, OC H_3), 3.80–3.60 (m, 3 H, H-3,4 and 5), 3.54 (dd, 1 H, $J_{1,2}$ 5.4, $J_{2,3}$ 7.1 Hz, H-2), 3.38 (dd, 1 H, $J_{4,5'}$ 2.2, $J_{5,5'}$ 12.5 Hz, H-5'), 2.2 (bs, 1 H, OH). For 14 β , ¹H NMR: δ 7.40–7.16 (m, 15 H, Ph), 5.80 (d, 1 H, J 8.7 Hz, NH), 5.10 (s, 2 H, PhC H_2 OCO), 4.80 (d, 1 H, $J_{1,2}$ 3.6 Hz, H-1), 4.67, 4.53 (2 d, 2 H, J 12.5 Hz, PhC H_2), 4.60–4.52 (m, 3 H, J 11.5 Hz, PhC H_2 , CH $_2$ CH), 4.09, 3.92 (2 dd, 2 H, J 10.8, $J_{\text{CH',CH}}$ 4.5 Hz, $J_{\text{CH',CH}}$ 2.1 Hz, C H_2 CH), 3.90–3.83 (m, 2 H, H-2,4), 3.70–3.56 (m, 6 H, H-3,5,5', and OCH $_3$).

Acetylation of 14α and 14β furnished 15α and 15β , respectively, as syrups. For 15α , $[\alpha]_D + 25.7^\circ$ (c 2.2, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 15 H, Ph), 5.65 (d, 1 H, J 8.2 Hz, NH), 5.18 (s, 2 H, PhC H_2 OCO), 5.02–4.97 (m, 1 H, H-3), 4.70, 4.50 (2 d, J 11.3 Hz, PhC H_2), 4.65–4.52 (m, 3 H, J 12.1 Hz, PhC H_2 , CH₂CH), 4.40 (d, 1 H, $J_{1,2}$ 5.1 Hz, H-1), 4.30–4.25 (m, 1 H, one proton of C H_2 CH), 4.09–4.01 (m, 1 H, one proton of C H_2 CH), 3.90–3.80 (m, 2 H, H-2,4), 3.74 (s, 3 H, OC H_3), 3.70–3.60 (m, 1 H, $J_{5,5'}$ 10.4 Hz, H-5), 3.47–3.40 (m, 1 H, $J_{5,5'}$ 10.4 Hz, H-5'), 2.01 (s, 3 H, C H_3 CO). Anal. Calcd for C₃₃H₃₇NO₁₀: C, 65.24; H, 6.10. Found: C, 64.92; H, 6.14. For 15β , $[\alpha]_D + 60.8^\circ$ (c 2.0, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 15 H, Ph), 5.82 (d, 1 H, J 8.9 Hz, NH), 5.11 (s, 2 H, PHC H_2 CO), 5.08 (dd, 1 H, $J_{2,3}$ 11.8, $J_{3,4}$ 3.9 Hz, H-3), 4.80 (d, 1 H, $J_{1,2}$ 4.0 Hz, H-1), 4.61, 4.55 (2 d, 2 H, J 11.9 Hz, PhC H_2), 4.59, 4.58 (2 d, 2 H, J 12.0 Hz, PhC H_2), 4.54–4.46 (m, 1 H, CH₂CH), 4.03, 3.97 (2 dd, 2 H, J 9.1, JCH',CH 2.5 Hz, JCH",CH 2.8 Hz, CH2CH), 3.88–3.78 (m, 2 H, H-2,4), 3.66 (s, 3 H, OC H_3), 3.65–3.62 (m, 2 H, H-5,5'), 2.02 (s, 3 H, C H_3 CO).

O-(2,4,6-Tri-O-benzyl-α- (20 α) and -β-D-galactopyranosyl)-N-benzyloxycarbonyl-serine methyl ester (20 β).—Assignment of compound 20 α (106.3 mg, 67%) and 20 β (33.2 mg, 20.9%) by ¹H NMR was difficult. Therefore, acetylation of 20 α, 20 β with Ac₂O in pyridine gave 21 α, 21 β, respectively. For 21 α, $[\alpha]_D$ +51° (c 6.7, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 20 H, Ph), 6.10 (d, 1 H, J 8.7 Hz, NH), 5.14 (dd, 1 H, J_{2,3} 10.4, J_{3,4} 3.4 Hz, H-3), 5.07 (s, 2 H, PhCH₂OCO), 4.79 (d, 1 H J_{1,2} 3.7 Hz, H-1), 4.80, 4.52 (2 d, 2 H, J 12.0 Hz, PhCH₂), 4.61, 4.55 (2 d, 2 H, J 11.6 Hz, PhCH₂), 4.57–4.44 (m, 3 H, J 12.1 Hz, PhCH₂), 4.61, 4.55 (2 d, 2 H, J 11.6 Hz, PhCH₂), 4.57–4.44 (m, 3 Hz, CHCH₂), 4.09–4.00 (m, 2 H, J_{3,4} 3.4 Hz, H-4, H-5), 3.98 (dd, 1 H, J_{1,2} 3.7, J_{2,3} 10.4 Hz, H-2), 3.68 (s, 3 H, OCH₃), 3.55–3.45 (m, 2 H, J_{5,6}′ 6.6, J_{5,6} 6.9 Hz, J_{6,6}′ 9.2 Hz, H-6,6′), 1.99 (s, 3 H, COCH₃). Anal. Calcd for C₄₁H₄₅NO₁₁: C, 67.68; H, 6.19. Found: C, 68.01; H, 6.27.

For **21** β , ¹H NMR: δ 7.38–7.15 (m, 20 H, Ph), 5.75 (d, 1 H, J 8.4 Hz, NH), 5.07 (s, 2 H, PhC H_2 OCO), 4.85 (dd, 1 H, $J_{2,3}$ 10.1, $J_{3,4}$ 4.2 Hz, H-3), 4.50–4.45 (m, 3 H, J 11.3 Hz, PhC H_2 , CH $_2$ CH), 4.43, 4.37 (2 d, 2 H, J 12.3 Hz, PhC H_2), 4.41, 4.38 (2 d, 2 H, J 10.6 Hz, PhC H_2), 4.37 (d, 1 H, $J_{1,2}$ 8.4 Hz, H-1), 4.16, 3.75 (2 dd, 2 H, J 10.2, $J_{\text{CH',CH}}$ 5.9, $J_{\text{CH',CH}}$ 3.0 Hz, CHC H_2), 4.14–4.00 (m, 2 H, $J_{3,4}$ 4.2 Hz, H-4,5), 3.93 (dd, 1 H, $J_{1,2}$ 8.4, $J_{2,3}$ 10.1 Hz, H-2), 3.75 (s, 3 H, OC H_3), 3.58–3.40 (m, 2 H, $J_{5,6'}$ 4.3, $J_{5,6'}$ 7.2 Hz, $J_{6,6'}$ 10.1 Hz, H-6,6'), 1.90 (s, 3 H, COC H_3).

O-(2,4,6-Tri-O-benzyl- α - (22α) and - β -D-galactopyranosyl)-N-tosyl-L-serine methyl ester (22β) .—For 22α (95 mg, 58.4%), $[\alpha]_D + 38^\circ$ (c 4.2, CHCl₃); ¹H NMR: δ 7.70

(d, 2 H, Ph of Ts), 7.40–7.10 (m, 17 H, Ph), 6.10 (d, 1 H, J 8.4 Hz, NH), 4.78, 4.58 (2 d, 2 H, J 11.9 Hz, PhC H_2), 4.71 (d, 1 H, $J_{1,2}$ 3.4 Hz, H-1), 4.60 (s, 2 H, PhC H_2), 4.51, 4.45 (2 d, 2 H, J 10.8 Hz, PhC H_2), 3.50 (s, 3 H, OC H_3), 2.42 (s, 3 H, PhC H_3). Anal. Calcd for C₃₈H₄₃NO₁₀S · 0.5H₂O: C, 63.86; H, 6.16. Found: C, 64.03; H, 6.09. For **22** β (35.6 mg, 21.8%), ¹H NMR: δ 7.60 (d, 2 H, Ph of Ts), 7.40–7.08 (m, 17 H, Ph), 5.65 (d, 1 H, J 8.8 Hz, NH), 4.79, 4.61 (2 d, 2 H, J 12.3 Hz, PhC H_2), 4.64, 4.63 (2 d, 2 H, J 12.4 Hz, PhC H_2), 4.28 (d, 1 H, J_{1,2} 8.4 Hz, H-1), 3.60 (s, 2 H, OC H_3), 2.40 (s, 3 H, PhC H_3).

Acetylation of **22** α gave **23** α ; $[\alpha]_D + 29^\circ$ (c 2.2, CHCl₃); ¹H NMR: δ 8.95 (d, 2 H, Ph of Ts), 7.40–7.20 (m, 17 H, Ph), 5.33–5.25 (m, 1 H, H-3), 4.90 (d, 1 H, $J_{1,2}$ 2.9 Hz, H-1), 4.68–4.40 (m, 7 H, 3 PhC H_2 , CH₂CH), 3.71 (s, 3 H, OC H_3), 2.40 (s, 3 H, PhC H_3), 2.20 (s, 3 H, NCOC H_3), 1.98 (s, 3 H, OCOC H_3). Acetylation of **22** β gave **23** β ; ¹H NMR: δ 8.94 (d, 2 H, Ph of Ts), 7.40–7.25 (m, 17 H, Ph), 5.30–5.20 (m, 1 H, H-3), 4.60–4.45 (m, 7 H, 3 PhC H_2 , CH₂CH), 4.40 (d, 1 H, $J_{1,2}$ 8.7 Hz, H-1), 3.68 (s, 3 H, OCC H_3), 2.44 (s, 3 H, PhC H_3), 2.22 (s, 3 H, NCOC H_3), 1.98 (s, 3 H, OCOC H_3).

O-(2,4,6-Tri-O-benzyl-α- (25α) and -β-D-mannopyranosyl)-N-benzyloxycarbonyl-L-serine methyl ester (25β).—For 25α (122.9 mg, 77.5%), [α]_D – 19° (c 3.7, CHCl₃); ¹H NMR: δ 7.40–7.05 (m, 20 H, Ph), 5.75 (d, 1 H, J 9.1 Hz, NH), 5.09 (s, 2 H, PhC H_2 OCO), 4.85 (d, 1 H, $J_{1,2}$ 1.3 Hz, H-1), 4.79, 4.46 (2 d, 2 H, J 10.8 Hz, PhC H_2), 4.71, 4.55 (2 d, 2 H, J 11.6 Hz, PhC H_2), 4.64, 4.48 (2 d, 2 H, J 11.8 Hz, PhC H_2), 4.53–4.45 (m, 1 H, OCH₂CH), 3.96–3.80 (m, 2 H, CHC H_2), 3.73 (s, 3 H, OC H_3), 3.72–3.58 (m, 6 H, H-2,3,4,5,6, and 6′), 2.17 (bs, 1 H, OH). Anal. Calcd for C₃₉H₄₃NO₁₀: C, 68.32; H, 6.28. Found: C, 68.21; H, 6.39. For 25 β (24.6 mg, 15.5%), [α]_D +11° (c 2.1, CHCl₃); ¹H NMR: δ 7.40–7.20 (m, 20 H, Ph), 5.72 (d, 1 H, J 8.6 Hz, NH), 5.12 (s, 2 H, PhC H_2 OCO), 4.91, 51 (2 d, 2 H, J 11.7 Hz, PhC H_2), 4.84, 4.55 (2 d, 2 H, J 10.8 Hz, PhC H_2), 4.64, 4.54 (2 d, 2 H, J 12.0 Hz, PhC H_2), 4.53–4.45 (m, 1 H, OCH₂CH), 4.47 (bs, 1 H, H-1), 4.35, 3.97 (2 dd, 2 H, J 10.2, JCH',CH 3.8, JCH",CH 7.1 Hz, CHC H_2), 3.80–3.40 (m, 6 H, H-2,3,4,5,6, and 6′), 3.61 (s, 3 H, OC H_3), 1.97 (bs, 1 H, OH).

Acetylation of **25** α gave **26** α as a syrup; $[\alpha]_D$ -25° (c 2.2, CHCl₃); ¹H NMR: δ 7.38-7.10 (m, 20 H, Ph), 5.87 (d, 1 H, J 8.5 Hz, NH), 5.19 (dd, 1 H, J_{2,3} 3.2, J_{3,4} 8.8 Hz, H-3), 5.10 (s, 2 H, PhCH₂OCO), 4.80 (d, 1 H, J_{1,2} 2.2 Hz, H-1), 4.64-4.40 (m, 7 H, 3 PhCH₂, CH₂CH), 4.10-3.72 (m, 5 H, OCHCH₂, H-2,4, and 5), 3.68 (s, 3 H, OCH₃), 3.67-3.58 (m, 2 H, H-6,6'), 1.93 (s, 3 H, COCH₃).

Acetylation of **25** β gave **26** β as a syrup; [α]_D + 14° (c 4.9, CHCl₃); ¹H NMR: δ 7.40–7.10 (m, 20 H, Ph), 5.68 (d, 1 H, J 8.9 Hz, NH), 5.33 (dd, 1 H, J_{2,3} 2.6, J_{3,4} 6.8 Hz, H-3), 5.12 (s, 2 H, PhCH₂OCO), 4.80–4.50 (m, 6 H, 3 PhCH₂), 4.42 (s, 1 H, H-1), 4.35 (dd, 1 H, J 10.2, J_{CH',CH} 2.7 Hz, one proton of OCHCH₂), 3.98–3.90 (m, 2 H, H-4, another proton of OCHCH₂), 3.80–3.70 (m, 2 H, H-2,5), 3.65 (s, 3 H, OCH₃), 3.50–3.35 (m, 2 H, H-6,6'), 1.90 (s, 3 H, OCOCH₃).

O-(2,4,6-Tri-O-benzyl- α - (27α) and- β -D-mannopyranosyl)-N-tosyl-L-serine methyl ester (27 β).—For 27α (102 mg, 62.5%); ¹H NMR: δ 7.69 (d, 2 H, Ph of Ts), 7.35–7.15 (m, 17 H, Ph), 5.68 (d, 1 H, J 8.9 Hz, NH), 4.81 (d, 1 H, J_{1,2} 1.6 Hz, H-1), 4.73, 4.50 (2 d, 2 H, J 11.4 Hz, PhCH₂), 4.70, 4.63 (2 d, 2 H, J 10.6 Hz, PhCH₂), 4.58, 4.52 (2 d, 2H, J 11.6 Hz, PhCH₂), 4.40–4.15 (m, 2 H, CH₂CH and H-4),

3.85–3.60 (m, 5 H, H-2,3,5,6 and 6'),3.50 (s, 3 H, OC H_3), 2.38 (s, 3 H, PhC H_3), 2.20 (bs, 1 H, OH). Anal. Calcd for C₃₈H₄₃NO₁₀S · 0.5H₂O: C, 63.86; H, 6.16. Found: C, 64.10; H, 6.21.

For 27β (41 mg, 25.1%), ¹H NMR: δ 7.73 (d, 2 H, Ph of Ts), 7.40–7.15 (m, 17 H, Ph), 5.70 (d, 1 H, J 8.6 Hz, NH), 4.88–4.45 (m, 7 H, 3 PhC H_2 , OCH $_2$ CH), 4.38 (d, 1 H, $J_{1,2}$ 1.7 Hz, H-1), 4.25 (dd, 1 H, J 11.0, $J_{\text{CH',CH}}$ 5.0 Hz, one proton of OCHC H_2), 4.15–4.08 (m, 2 H, H-4 and another proton of OCHC H_2), 3.80–3.55 (m, 5 H, H-2,3,5,6, and 6'), 3.48 (s, 3 H, OC H_3), 2.37 (s, 3 H, PhC H_3), 1.70 (bs, 1 H, OH).

Acetylation of 27α gave 28α as a syrup in a quantitative yield; $[\alpha]_D - 48^\circ$ (c 2.3, CHCl₃); ¹H NMR: δ 7.90 (d, 2 H, Ph of Ts), 7.40–7.15 (m, 17 H, Ph), 5.20 (dd, 1 H, $J_{2,3}$ 3.7, $J_{3,4}$ 8.8 Hz, H-3), 4.90 (d, 1 H, J 1.7 Hz, H-1), 4.78–4.40 (m, 7 H, 3 PhC H_2 and OCH₂CH), 4.18–3.70 (m, 7 H, CHC H_2 , H-2,4,5,6, and 6'), 3.60 (s, 3 H, OC H_3), 2.40 (s, 3 H, PhC H_3), 2.21 (s, 3 H, NCOC H_3), 1.95 (s, 3 H, OCOC H_3). Anal. Calcd for C₁₂H₁₇NO₁₂S: C, 63.88; H, 5.96; Found: C, 64.05; H, 6.11.

Acetylation of **27** β gave **28** β as a syrup in a quantitative yield; $[\alpha]_D - 7.0^\circ$ (c 2.5, CHCl₃); ¹H NMR: δ 8.01 (d, 2 H, Ph of Ts), 7.40–7.10 (m, 17 H, Ph), 5.43 (dd, 1 H, $J_{2,3}$ 3.6, $J_{3,4}$ 5.4 Hz, H-3), 4.90–4.60 (m, 7 H, 3 PhC H_2 and OCH₂CH), 4.58 (bs, 1 H, H-1), 4.45, 4.07 (2dd, 2 H, J 9.8, $J_{CH',CH}$ 4.7, $J_{CH',CH}$ 6.1 Hz, CHC H_2), 4.32–4.25 (m, 1 H, $J_{3,4}$ 5.0, $J_{4,5}$ 5.3 Hz, H-4), 3.82–3.67 (m, 2 H, H-2,5), 3.65 (s, 3 H, OC H_3), 3.60–3.43 (m, 2 H, H-6,6'), 2.30 (s, 3 H, PhC H_3), 2.19 (s, 3 H, NCOC H_3), 1.85 (s, 3 H, OCOC H_3).

O-(2,4,6-Tri-O-benzyl-D-mannopyranosyl)-N-benzyloxycarbonyl-L-threonine methyl ester (29).—Compound 29 was obtained as an α , β mixture (126 mg, 78%, α : β 3:1); [α]_D +9.4° (c 2.7, CHCl₃); ¹H NMR: δ 7.48–7.15 (m, 20 H, Ph), 5.48 (d, 0.25 H, J 8.3 Hz, NH of β anomer), 5.37 (d, 0.75 H, J 8.4 Hz, NH of α anomer), 5.12 (s, 1.5 H, PhC H_2 OCO of α anomer), 5.11 (s, 0.5 H, PhC H_2 OCO of β anomer), 4.91 (d, 0.75 H, $J_{1,2}$ 1.7 Hz, H-1 of α anomer), 4.75–4.45 (m, 9 H, 3 PhC H_2 , H-2, OCHCH $_3$, CHNH), 4.36 (d, 0.25 H, $J_{1,2}$ 2.1 Hz, H-1 of β anomer), 3.70 (s, 2.25 H, OC H_3 of α anomer), 3.50 (s, 0.75 H, OC H_3 of β anomer), 2.25 (bs, 0.75 H, OH of α anomer), 1.61 (bs, 0.25 H, OH of β anomer), 1.35 (d, 2.25 H, J 4.3 Hz, OCHC H_3 of α anomer), 1.34 (d, 0.75 H, J 4.9 Hz, OCHC H_3 of β anomer). Anal. Calcd for C₄₀H₄₅NO₁₀: C, 68.67; H, 6.44. Found: C, 68.32; H, 6.51.

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